

Medical Coverage Policy | Identification of Microorganisms Using Nucleic Acid Probes



EFFECTIVE DATE: 01 | 01 | 2024

POLICY LAST REVIEWED: 09 | 06 | 2023

OVERVIEW

Nucleic acid probes are available for the identification of a wide variety of microorganisms. Nucleic acid probes can also be used to quantitate the number of microorganisms present. This technology offers advantages over standard techniques when rapid identification is clinically important, microbial identification using standard culture is difficult or impossible, and/or treatment decisions are based on quantitative results.

The following test(s) are addressed in this policy:

- MicroGenDX qPCR & NGS For Infection (MicroGenDX) CPT code 0112U
- ePlex BCID Fungal Pathogens Panel (GenMark Diagnostics, Inc.) CPT code 0140U
- ePlex BCID Gram-Positive Panel (GenMark Diagnostics, Inc.) CPT code 0141U
- ePlex BCID Gram-Negative Panel (GenMark Diagnostics, Inc.) CPT code 0142U

MEDICAL CRITERIA

Medicare Advantage Plans and Commercial Products

MicroGenDX qPCR & NGS For Infection (CPT code 0112U), ePlex BCID Fungal Pathogens Panel (CPT code 0140U), ePlex BCID Gram-Positive Panel (CPT code 0141U) and ePlex BCID Gram-Negative Panel (CPT code 0142U)

The use of nucleic acid testing using a direct or amplified probe technique (without quantification of viral load) may be considered medically necessary for the following microorganisms:

- *Bartonella henselae* or *quintana*
- *Bordetella pertussis*
- *Candida* species
- *Chlamydia pneumoniae*
- *Chlamydia trachomatis*
- *Clostridium difficile*
- *Enterococcus*, vancomycin-resistant (e.g., *enterococcus vanA*, *vanB*)
- Enterovirus
- Herpes simplex virus
- Human papillomavirus
- Influenza virus
- *Legionella pneumophila*
- Mumps
- *Mycobacterium* species
- *Mycobacterium tuberculosis*
- *Mycobacterium avium-intracellulare*
- *Mycoplasma pneumoniae*
- *Neisseria gonorrhoeae*
- Rubeola (measles)
- Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2)
- *Staphylococcus aureus*
- *Staphylococcus aureus*, methicillin-resistant

- Streptococcus, group A
- Streptococcus, group B
- Trichomonas vaginalis
- Zika virus

The use of nucleic acid testing using a direct or amplified probe technique (with or without quantification of viral load) may be considered medically necessary for the following microorganisms:

- Cytomegalovirus
- Hepatitis B virus
- Hepatitis C virus
- HIV-1
- HIV-2
- Human herpesvirus 6

The use of the following nucleic acid testing panel (without quantification of viral load) may be considered medically necessary

- Respiratory virus panel

PRIOR AUTHORIZATION

Medicare Advantage Plans and Commercial Products

Prior authorization is required for the following Proprietary Laboratory Analyses (PLA) tests:

- MicroGenDX qPCR & NGS For Infection (MicroGenDX) CPT code 0112U
- ePlex BCID Fungal Pathogens Panel (GenMark Diagnostics, Inc.) CPT code 0140U
- ePlex BCID Gram-Positive Panel (GenMark Diagnostics, Inc.) CPT code 0141U
- ePlex BCID Gram-Negative Panel (GenMark Diagnostics, Inc.) CPT code 0142U

POLICY STATEMENT

Medicare Advantage Plans and Commercial Products

The following Proprietary Laboratory Analyses (PLA) tests may be considered medically necessary when the medical criteria above has been met:

- MicroGenDX qPCR & NGS For Infection (MicroGenDX) CPT code 0112U
- ePlex BCID Fungal Pathogens Panel (GenMark Diagnostics, Inc.) CPT code 0140U
- ePlex BCID Gram-Positive Panel (GenMark Diagnostics, Inc.) CPT code 0141U
- ePlex BCID Gram-Negative Panel (GenMark Diagnostics, Inc.) CPT code 0142U

The use of nucleic acid testing with quantification of viral load is considered for microorganisms that are not included in the list of microorganisms above for which probes with or without quantification are not covered for Medicare Advantage Plans and not medically necessary for Commercial Products as the evidence is insufficient to determine that the technology results in an improvement in the net health outcome.

The use of nucleic acid testing using a direct or amplified probe technique is considered for the following microorganisms not covered for Medicare Advantage Plans and not medically necessary for Commercial Products as the evidence is insufficient to determine that the technology results in an improvement in the net health outcome:

- Gardnerella vaginalis
- Hepatitis G

The use of the following nucleic acid testing panels (with or without quantification of viral load for viral panel elements) is not covered for Medicare Advantage Plans and not medically necessary for Commercial Products as the evidence is insufficient to determine that the technology results in an improvement in the net health outcome:

- Central nervous system pathogen panel
- Gastrointestinal pathogen panel

Note: Laboratories are not allowed to obtain clinical authorization or participate in the authorization process on behalf of the ordering physician. Only the ordering physician shall be involved in the authorization, appeal or other administrative processes related to prior authorization/medical necessity.

In no circumstance shall a laboratory or a physician/provider use a representative of a laboratory or anyone with a relationship to a laboratory and/or a third party to obtain authorization on behalf of the ordering physician, to facilitate any portion of the authorization process or any subsequent appeal of a claim where the authorization process was not followed and/or a denial for clinical appropriateness was issued, including any element of the preparation of necessary documentation of clinical appropriateness. If a laboratory or a third party is found to be supporting any portion of the authorization process, BCBSRI will deem the action a violation of this policy and severe action will be taken up to and including termination from the BCBSRI provider network. If a laboratory provides a laboratory service that has not been authorized, the service will be denied as the financial liability of the participating laboratory and may not be billed to the member.

Commercial Products

Some genetic testing services are not covered and a contract exclusion for any self-funded group that has excluded the expanded coverage of biomarker testing related to the state mandate, R.I.G.L. §27-19-81 described in the Biomarker Testing Mandate policy. For these groups, a list of which genetic testing services are covered with prior authorization, are not medically necessary or are not covered because they are a contract exclusion can be found in the Coding section of the Genetic Testing Services or Proprietary Laboratory Analyses policies. Please refer to the appropriate Benefit Booklet to determine whether the member's plan has customized benefit coverage. Please refer to the list of Related Policies for more information.

COVERAGE

Benefits may vary between groups and contracts. Please refer to the appropriate Benefit Booklet, Evidence of Coverage or Subscriber Agreement for applicable laboratory benefits/coverage.

BACKGROUND

Nucleic Acid Probes

A nucleic acid probe is used to detect and identify species or subspecies of organisms by identifying nucleic acid sequences in a sample. Nucleic acid probes detect genetic materials, such as RNA or DNA, unlike other tests, which use antigens or antibodies to diagnose organisms.

The availability of nucleic acid probes has permitted the rapid direct identification of microorganism DNA or RNA. Amplification techniques result in exponential increases in copy numbers of a targeted strand of microorganism-specific DNA. The most used amplification technique is polymerase chain reaction (PCR) or reverse transcriptase PCR. In addition to PCR, other nucleic acid amplification techniques have been developed, such as transcription-mediated amplification, loop-mediated isothermal DNA amplification, strand displacement amplification, nucleic acid sequence-based amplification, and branched-chain DNA signal amplification. After amplification, target DNA can be readily detected using a variety of techniques. The amplified product can also be quantified to assess how many microorganisms are present. Quantification of the number of nucleic acids permits serial assessments of response to treatment; the most common clinical application of quantification is the serial measurement of HIV RNA (called viral load).

The direct probe technique, amplified probe technique, and probe with quantification methods vary based on the degree to which the nucleic acid is amplified and the method for measurement of the signal. The direct probe technique refers to detection methods in which nucleic acids are detected without an initial amplification step. The amplified probe technique refers to detection methods in which either target, probe, or signal amplification is used to improve the sensitivity of the assay over direct probe techniques, without quantification of nucleic acid amounts.

- Target amplification methods include PCR (including PCR using specific probes, nested or multiplex PCR), nucleic acid-based sequence amplification, transcription-mediated amplification, and strand displacement amplification. Nucleic acid-based sequence amplification and transcription-mediated amplification involve amplification of an RNA (rather than a DNA) target.
- Probe amplification methods include ligase chain reaction.
- Signal amplification methods include branched DNA (bDNA) probes and hybrid capture methods using an anti-DNA/RNA hybrid antibody.

The probe with quantification techniques refers to quantitative PCR or real-time PCR methods that use a reporter at each stage of the PCR to generate absolute or relative amounts of a known nucleic acid sequence in the original sample. These methods may use DNA-specific dyes (ethidium bromide or SYBR green), hybridization probes (cleavage-based [TaqMan] or displacable), or primer incorporated probes.

Direct assays will generally have lower sensitivity than amplified probes. In practice, most commercially available probes are amplified, with a few exceptions. For this evidence review, indications for direct and/or amplified probes without quantification are considered together, while indications for a probe with quantification are considered separately.

Classically, identification of microorganisms relies either on the culture of body fluids or tissues or identification of antigens, using a variety of techniques including direct fluorescent antibody technique and qualitative or quantitative immunoassays. These techniques are problematic when the microorganism exists in very small numbers or is technically difficult to culture. Indirect identification of microorganisms by immunoassays for specific antibodies reactive with the microorganism is limited by difficulties in distinguishing between past exposure and current infection.

Potential reasons for a nucleic acid probe to be associated with improved clinical outcomes compared with standard detection techniques include the following (note: in all cases, for there to be clinical utility, making a diagnosis should be associated with changes in clinical management, which could include initiation of effective treatment, discontinuation of other therapies, or avoidance of invasive testing):

- Significantly improved speed and/or efficiency in making a diagnosis.
- Improved likelihood of obtaining any diagnosis in cases where standard culture is difficult. Potential reasons for difficulty in obtaining standard culture include low numbers of the organisms (e.g., HIV), fastidious or lengthy culture requirements (e.g., Mycobacteria, Chlamydia, Neisseria species), or difficulty in collecting an appropriate sample (e.g., herpes simplex encephalitis).
- There is no way to definitively make a diagnosis without nucleic acid testing.
- The use of nucleic acid probe testing provides qualitatively different information than that available from standard cultures, such as information regarding disease prognosis or response to treatment. These include cases where quantification of viral load provides prognostic information or is used to measure response to therapy.

The risks of nucleic acid testing include false-positive and false-negative results, inaccurate identification of pathogens by the device, inaccurate interpretation of test results, or incorrect operation of the instrument.

- False-positive results can lead to unnecessary treatment, with its associated toxicities and side effects, including allergic reaction. In addition, true diagnosis and treatment could be delayed or missed altogether.
- False-negative results could delay diagnosis and initiation of proper treatment.
- It is possible that these risks can be mitigated by the use of a panel of selected pathogens indicated by the clinical differential diagnosis while definitive culture results are pending.

For individuals who have signs and/or symptoms of meningitis and/or encephalitis who receive a nucleic acid-based central nervous system pathogen panel, the evidence includes a systematic review and a pivotal prospective study. Relevant outcomes include test accuracy and validity, other test performance measures, medication use, symptoms, and change in disease status. Access to a rapid method that can simultaneously

test for multiple pathogens may lead to the faster initiation of more effective treatment and conservation of cerebrospinal fluid. The available central nervous system panel is highly specific for the included organisms, but the sensitivity for each pathogen is not well-characterized. More than 15% of positives in the largest clinical validity study were false-positives. A negative panel result does not exclude infection due to pathogens not included in the panel. The evidence is insufficient to determine that the technology results in an improvement in the net health outcome.

For individuals who have signs and/or symptoms of gastroenteritis who receive a nucleic acid-based gastrointestinal pathogen panel, the evidence includes prospective and retrospective evaluations of the tests' sensitivity and specificity and prospective studies on utility. Relevant outcomes include test accuracy and validity, other test performance measures, medication use, symptoms, and change in disease status. The evidence suggests that pathogen panels are likely to identify both bacterial and viral pathogens with high sensitivity, compared with standard methods. Access to a rapid method for etiologic diagnosis of infections may lead to more effective early treatment and infection control measures. However, in most instances, when a specific pathogen is suspected, individual tests could be ordered. There may be a subset of patients with an unusual presentation who would warrant testing for a panel of pathogens at once, but that subset has not been well defined.

The evidence is insufficient to determine that the technology results in an improvement in the net health outcome. For individuals who have signs and/or symptoms of respiratory infection who receive a nucleic acid-based respiratory pathogen panel, the evidence includes a systematic review and 2 randomized controlled trials (RCTs). Relevant outcomes include test accuracy and validity, other test performance measures, medication use, symptoms, and change in disease status. The systematic review reported that all 3 reviewed multiplex polymerase chain reaction systems were highly accurate. One RCT and 1 quasi-RCT evaluated utility of a respiratory panel and found benefits in time-to-treat and length of hospital stay. In addition, 1 subanalysis found fewer antibiotics being prescribed for patients diagnosed with the panel. The panel did not significantly affect duration of antibiotic use, readmission, or mortality rates. The evidence is sufficient to determine that the technology results in an improvement in the net health outcome.

CODING

Medicare Advantage Plans and Commercial Products

The following CPT code(s) are covered for Medicare Advantage Plans and Commercial Products when medical criteria above are met:

This code can be used for MicroGenDX qPCR & NGS For Infection (MicroGenDX)

0112U Infectious agent detection and identification, targeted sequence analysis (16S and 18S rRNA genes) with drug-resistance gene

This code can be used for ePlex BCID Fungal Pathogens Panel (GenMark Diagnostics, Inc.)

0140U Infectious disease (fungi), fungal pathogen identification, DNA (15 fungal targets), blood culture, amplified probe technique, each target reported as detected or not detected

This code can be used for ePlex BCID Gram-Positive Panel (GenMark Diagnostics, Inc.)

0141U Infectious disease (bacteria and fungi), gram-positive organism identification and drug resistance element detection, DNA (20 gram-positive bacterial targets, 4 resistance genes, 1 pan gram-negative bacterial target, 1 pan Candida target), blood culture, amplified probe technique, each target reported as detected or not detected

This code can be used for ePlex BCID Gram-Negative Panel (GenMark Diagnostics, Inc.)

0142U Infectious disease (bacteria and fungi), gram-negative bacterial identification and drug resistance element detection, DNA (21 gram-negative bacterial targets, 6 resistance genes, 1 pan gram-positive bacterial target, 1 pan Candida target), amplified probe technique, each target reported as detected or not detected

RELATED POLICIES

Biomarker Testing Mandate
Proprietary Laboratory Analyses (PLA)

PUBLISHED

Provider Update, November 2023

REFERENCES

1. He T, Kaplan S, Kamboj M, et al. Laboratory Diagnosis of Central Nervous System Infection. *Curr Infect Dis Rep.* Nov 2016; 18(11): 35. PMID 27686677
2. Tansarli GS, Chapin KC. Diagnostic test accuracy of the BioFire® FilmArray® meningitis/encephalitis panel: a systematic review and meta-analysis. *Clin Microbiol Infect.* Mar 2020; 26(3): 281-290. PMID 31760115
3. Leber AL, Everhart K, Balada-Llasat JM, et al. Multicenter Evaluation of BioFire FilmArray Meningitis/Encephalitis Panel for Detection of Bacteria, Viruses, and Yeast in Cerebrospinal Fluid Specimens. *J Clin Microbiol.* Sep 2016; 54(9): 2251-61. PMID 27335149
4. Graf EH, Farquharson MV, Cárdenas AM. Comparative evaluation of the FilmArray meningitis/encephalitis molecular panel in a pediatric population. *Diagn Microbiol Infect Dis.* Jan 2017; 87(1): 92-94. PMID 27771208
5. Hanson KE, Slechta ES, Killpack JA, et al. Preclinical Assessment of a Fully Automated Multiplex PCR Panel for Detection of Central Nervous System Pathogens. *J Clin Microbiol.* Mar 2016; 54(3): 785-7. PMID 26719436
6. Gastrointestinal Tract Infections. https://www.uib.cat/depart/dba/microbiologia/ADSenfcomI/material_archivos/infeccion%20gastrointestinal.pdf. Accessed May 12, 2023.
7. Bintsis T. Foodborne pathogens. *AIMS Microbiol.* 2017; 3(3): 529-563. PMID 31294175
8. Sattar SBA, Singh S. Bacterial Gastroenteritis. [Updated 2021 Aug 11]. In: StatPearls [Internet]. Treasure Island, FL: StatPearls Publishing; 2019 Jan. <https://www.ncbi.nlm.nih.gov/books/NBK513295/>. Accessed May 12, 2023.
9. Burden of Norovirus Illness in the U.S. Centers for Disease Control and Prevention. <https://www.cdc.gov/norovirus/trends-outbreaks/burden-US.html>. Last reviewed March 5, 2021. Accessed May 12, 2023.
10. Centers for Medicare & Medicaid Coverage. Local Coverage Determination (LCD): Foodborne Gastrointestinal Panels Identified by Multiplex Nucleic Acid Amplification (NAATs) (L37709). CMS.gov. <https://www.cms.gov/medicare-coverage-database/view/lcd.aspx?lcdid=37709&ver=20&bc=0>. Revised April 17, 2022. Accessed May 11, 2023.
11. Beckmann C, Heining U, Marti H, et al. Gastrointestinal pathogens detected by multiplex nucleic acid amplification testing in stools of pediatric patients and patients returning from the tropics. *Infection.* Dec 2014; 42(6): 961-70. PMID 25015433
12. Borst A, Box AT, Fluit AC. False-positive results and contamination in nucleic acid amplification assays: suggestions for a prevent and destroy strategy. *Eur J Clin Microbiol Infect Dis.* Apr 2004; 23(4): 289-99. PMID 15015033
13. Evaluation of automatic class III designation (de novo) for xTAG gastrointestinal pathogen panel (GPP) decision summary. Food and Drug Administration. https://www.accessdata.fda.gov/cdrh_docs/reviews/K121454.pdf. Accessed May 11, 2023.
14. Claas EC, Burnham CA, Mazzulli T, et al. Performance of the xTAG® gastrointestinal pathogen panel, a multiplex molecular assay for simultaneous detection of bacterial, viral, and parasitic causes of infectious gastroenteritis. *J Microbiol Biotechnol.* 2013; 23(7): 1041-5. PMID 23711521
15. Khare R, Espy MJ, Cebelski E, et al. Comparative evaluation of two commercial multiplex panels for detection of gastrointestinal pathogens by use of clinical stool specimens. *J Clin Microbiol.* Oct 2014; 52(10): 3667-73. PMID 25100818
16. Buchan BW, Olson WJ, Pezewski M, et al. Clinical evaluation of a real-time PCR assay for identification of Salmonella, Shigella, Campylobacter (Campylobacter jejuni and C. coli), and shiga toxin-producing Escherichia coli isolates in stool specimens. *J Clin Microbiol.* Dec 2013; 51(12): 4001-7. PMID 24048539

17. Al-Talib H, Latif B, Mohd-Zain Z. Pentaplex PCR assay for detection of hemorrhagic bacteria from stoolsamples. *J Clin Microbiol.* Sep 2014; 52(9): 3244-9. PMID 24958797
18. Jiang Y, Fang L, Shi X, et al. Simultaneous detection of five enteric viruses associated withgastroenteritis by use of a PCR assay: a single real-time multiplex reaction and its clinical application. *J Clin Microbiol.* Apr 2014; 52(4): 1266-8. PMID 24478418
19. Freeman K, Mistry H, Tsertsvadze A, et al. Multiplex tests to identify gastrointestinal bacteria, virusesand parasites in people with suspected infectious gastroenteritis: a systematic review and economicanalysis. *Health Technol Assess.* Apr 2017; 21(23): 1-188. PMID 28619124
20. Kosai K, Suzuki H, Tamai K, et al. Multicenter evaluation of Verigene Enteric Pathogens Nucleic AcidTest for detection of gastrointestinal pathogens. *Sci Rep.* Feb 04 2021; 11(1): 3033. PMID 33542335
21. Meltzer AC, Newton S, Lange J, et al. A randomized control trial of a multiplex gastrointestinal PCRpanel versus usual testing to assess antibiotics use for patients with infectious diarrhea in theemergency department. *J Am Coll Emerg Physicians Open.* Feb 2022; 3(1): e12616. PMID 35072157
22. Cybulski RJ, Bateman AC, Bourassa L, et al. Clinical Impact of a Multiplex Gastrointestinal PolymeraseChain Reaction Panel in Patients With Acute Gastroenteritis. *Clin Infect Dis.* Nov 13 2018; 67(11): 1688-1696. PMID 29697761
23. Beal SG, Tremblay EE, Toffel S, et al. A Gastrointestinal PCR Panel Improves Clinical Management andLowers Health Care Costs. *J Clin Microbiol.* Jan 2018; 56(1). PMID 29093106
24. Darie AM, Khanna N, Jahn K, et al. Fast multiplex bacterial PCR of bronchoalveolar lavage for antibioticstewardship in hospitalised patients with pneumonia at risk of Gram-negative bacterial infection(Flagship II): a multicentre, randomised controlled trial. *Lancet Respir Med.* Sep 2022; 10(9): 877-887.PMID 35617987
25. Clark TW, Lindsley K, Wigmosta TB, et al. Rapid multiplex PCR for respiratory viruses reduces time toresult and improves clinical care: Results of a systematic review and meta-analysis. *J Infect.* May 2023;86(5): 462-475. PMID 36906153
26. Huang HS, Tsai CL, Chang J, et al. Multiplex PCR system for the rapid diagnosis of respiratory virusinfection: systematic review and meta-analysis. *Clin Microbiol Infect.* Oct 2018; 24(10): 1055-1063.PMID 29208560
27. Mansuy JM, Mengelle C, Da Silva I, et al. Performance of a rapid molecular multiplex assay for thedetection of influenza and picornaviruses. *Scand J Infect Dis.* Dec 2012; 44(12): 963-8. PMID 22830610
28. Dabisch-Ruthe M, Vollmer T, Adams O, et al. Comparison of three multiplex PCR assays for thedetection of respiratory viral infections: evaluation of xTAG respiratory virus panel fast assay,RespiFinder 19 assay and RespiFinder SMART 22 assay. *BMC Infect Dis.* Jul 24 2012; 12: 163. PMID22828244
29. Pierce VM, Hodinka RL. Comparison of the GenMark Diagnostics eSensor respiratory viral panel toreal-time PCR for detection of respiratory viruses in children. *J Clin Microbiol.* Nov 2012; 50(11): 3458-65. PMID 22875893
30. Andrews D, Chetty Y, Cooper BS, et al. Multiplex PCR point of care testing versus routine, laboratory-based testing in the treatment of adults with respiratory tract infections: a quasi-randomised studyassessing impact on length of stay and antimicrobial use. *BMC Infect Dis.* Oct 10 2017; 17(1): 671.PMID 29017451
31. Brendish NJ, Malachira AK, Armstrong L, et al. Routine molecular point-of-care testing for respiratoryviruses in adults presenting to hospital with acute respiratory illness (ResPOC): a pragmatic, open-label,randomised controlled trial. *Lancet Respir Med.* May 2017; 5(5): 401-411. PMID 28392237
32. Cytomegalovirus (CMV) and Congenital CMV Infection: Laboratory Testing. Centers for Disease Controland Prevention. <https://www.cdc.gov/cmV/clinical/lab-tests.html>. Page last reviewed April 28, 2020.Accessed May 12, 2023.
33. Mycoplasma pneumoniae Infections: Diagnostic Methods. Center for Disease Control and Prevention.<https://www.cdc.gov/pneumonia/atypical/mycoplasma/hcp/diagnostic-methods.html>. Last ReviewedJune 5, 2020. Accessed May 12, 2023.
34. Zika Virus: Testing Guidance. Center for Disease Control and Prevention. <https://www.cdc.gov/zika/hc-providers/testing-guidance.html>. Last Reviewed December 9, 2019. Accessed May 12, 2023.
35. MacCannell T, Umscheil CA, Agarwal RK, et al. Guideline for the Prevention and Control of NorovirusGastroenteritis Outbreaks in Healthcare Settings. CDC. Updated February 15,

- 2017.<https://www.cdc.gov/infectioncontrol/pdf/guidelines/norovirus-guidelines.pdf>. Accessed May 12, 2023.
36. Hall AJ, Vinje J, Lopman B, et al. Updated Norovirus Outbreak Management and Disease Prevention Guidelines. CDC MMWR. Published March 4, 2011. <https://www.cdc.gov/mmwr/pdf/rr/rr6003.pdf>. Accessed May 12, 2023.
37. Workowski KA, Bolan GA, Workowski KA, et al. Sexually transmitted diseases treatment guidelines, 2015. MMWR Recomm Rep. Jun 05 2015; 64(RR-03): 1-137. PMID 26042815
38. Sexually Transmitted Infections Treatment Guidelines, 2021. Center for Disease Control and Prevention. <https://www.cdc.gov/std/treatment-guidelines/default.htm>. Last Reviewed July 22, 2021. Accessed May 12, 2023.
39. Recommendations for the Laboratory-Based Detection of Chlamydia trachomatis and Neisseria gonorrhoeae 2014. CDC MMWR. Published March 14, 2014. <https://www.cdc.gov/mmwr/preview/mmwrhtml/rr6302a1.htm>. Accessed May 12, 2023.
40. Updated Guidelines for the Use of Nucleic Acid Amplification Tests in the Diagnosis of Tuberculosis. CDC MMWR. Published January 16, 2009. https://www.cdc.gov/mmwr/preview/mmwrhtml/mm5801a3.htm?s_cid=mm5801a3_e. Accessed May 12, 2023.
41. NIH Guidelines for the Prevention and Treatment of Opportunistic Infections in Adults and Adolescents with HIV. Updated April 12, 2022. <https://clinicalinfo.hiv.gov/en/guidelines/adult-and-adolescent-opportunistic-infection/whats-new-guidelines>. Accessed on May 12, 2023.
42. Miller JM, Binnicker MJ, Campbell S, et al. A Guide to Utilization of the Microbiology Laboratory for Diagnosis of Infectious Diseases: 2018 Update by the Infectious Diseases Society of America and the American Society for Microbiology. Clin Infect Dis. Aug 31 2018; 67(6): e1-e94. PMID 29955859
43. Tunkel AR, Hasbun R, Bhimraj A, et al. 2017 Infectious Diseases Society of America's Clinical Practice Guidelines for Healthcare-Associated Ventriculitis and Meningitis. Clin Infect Dis. Mar 15 2017; 64(6): e34-e65. PMID 28203777
44. Tunkel AR, Glaser CA, Bloch KC, et al. The management of encephalitis: clinical practice guidelines by the Infectious Diseases Society of America. Clin Infect Dis. Aug 01 2008; 47(3): 303-27. PMID 18582201
45. Lee DH, Vielemeyer O. Analysis of overall level of evidence behind Infectious Diseases Society of America practice guidelines. Arch Intern Med. Jan 10 2011; 171(1): 18-22. PMID 21220656
46. McDonald LC, Gerding DN, Johnson S, et al. Clinical Practice Guidelines for Clostridium difficile Infection in Adults and Children: 2017 Update by the Infectious Diseases Society of America (IDSA) and Society for Healthcare Epidemiology of America (SHEA). Clin Infect Dis. Mar 19 2018; 66(7): e1-e48. PMID 29462280
47. Shane AL, Mody RK, Crump JA, et al. 2017 Infectious Diseases Society of America Clinical Practice Guidelines for the Diagnosis and Management of Infectious Diarrhea. Clin Infect Dis. Nov 29 2017; 65(12): e45-e80. PMID 29053792
48. Pappas PG, Kauffman CA, Andes DR, et al. Clinical Practice Guideline for the Management of Candidiasis: 2016 Update by the Infectious Diseases Society of America. Clin Infect Dis. Feb 15 2016; 62(4): e1-50. PMID 26679628
49. Infectious Diseases Society of America Guidelines on the Diagnosis of COVID-19. Published May 6, 2020. Updated December 23, 2020. <https://www.idsociety.org/practice-guideline/covid-19-guideline-diagnostics/>. Accessed on May 12, 2023.
50. Nellore A, Huprikar S. Vancomycin-resistant Enterococcus in solid organ transplant recipients: Guidelines from the American Society of Transplantation Infectious Diseases Community of Practice. Clin Transplant. Sep 2019; 33(9): e13549. PMID 30913322
51. Kimberlin DW, Barnett ED, Lynfield R, et al. Red Book: 2021 Report on the Committee on Infectious Diseases, 32nd Edition. American Academy of Pediatrics: 2021.
52. Puopolo KM, Lynfield R, Cummings JJ, et al. Management of Infants at Risk for Group B Streptococcal Disease. Pediatrics. Aug 2019; 144(2). PMID 31285392
53. Riddle MS, DuPont HL, Connor BA. ACG Clinical Guideline: Diagnosis, Treatment, and Prevention of Acute Diarrheal Infections in Adults. Am J Gastroenterol. May 2016; 111(5): 602-22. PMID 27068718
54. Filkins L, Hauser J, Robinson-Dunn B, Tibbetts R, Boyanton B, Revell P. Guidelines for the Detection and Identification of Group B Streptococcus. American Society for Microbiology. Published March

10,2020. Updated July 23, 2021. <https://asm.org/Guideline/Guidelines-for-the-Detection-and-Identification-of>. Accessed May 12, 2023.

CLICK THE ENVELOPE ICON BELOW TO SUBMIT COMMENTS

This medical policy is made available to you for informational purposes only. It is not a guarantee of payment or a substitute for your medical judgment in the treatment of your patients. Benefits and eligibility are determined by the member's subscriber agreement or member certificate and/or the employer agreement, and those documents will supersede the provisions of this medical policy. For information on member-specific benefits, call the provider call center. If you provide services to a member which are determined to not be medically necessary (or in some cases medically necessary services which are non-covered benefits), you may not charge the member for the services unless you have informed the member and they have agreed in writing in advance to continue with the treatment at their own expense. Please refer to your participation agreement(s) for the applicable provisions. This policy is current at the time of publication; however, medical practices, technology, and knowledge are constantly changing. BCBSRI reserves the right to review and revise this policy for any reason and at any time, with or without notice. Blue Cross & Blue Shield of Rhode Island is an independent licensee of the Blue Cross and Blue Shield Association.

